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PATENT

Attorney Docket No. DHI-06207

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of: Yung T. Huang
Serial No.: 09/844,311
Filed: 04/27/01
Entitled: **CELLS FOR DETECTION OF
ENTEROVIRUSES**

Group No.: 1648
Examiner: Sharon A. Foley

**DECLARATION UNDER 1.132 BY
DR. YUNG T. HUANG**

Mail Stop - AF
Commissioner for Patents
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CERTIFICATE OF TRANSMISSION UNDER 37 C.F.R. § 1.8(a)(1)(i)(B)

I hereby certify that this correspondence (along with any documents referred to as being attached or enclosed) is, on the date shown below, being transmitted by facsimile to the Patent and Trademark Office.

Date:

3/2/2004

By:

Cliff Cannon-Clin
Cliff Cannon-Clin

Madam:

1. I, Yung T. Huang, am inventor of the pending claims in the instant application, and am the subject of the attached Curriculum Vitae (Tab 1) and author of the publications shown on the list attached thereto. On the basis of the information and facts contained in these documents, I submit that I am qualified to speak on the level of ordinary skill in the art of the claimed invention.

2. The Examiner rejected Claims 1-4 and 6-14 as being allegedly obvious over Scholl *et al.*¹ and Powell *et al.*,² and also rejected Claim 5 under 35 U.S.C. §103(a) for

¹ US patent 6,168,915.

² Powell *et al.* (1998) J. Gen. Virol. 79:1707-1713.

³ Paper No. 17, middle of page 3.

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alleged obviousness over Scholl *et al.*, Powell *et al.*, Spiller *et al.*⁴, and either the sequence alignment of SEQ ID NO:1 with GenEmbl accession no. M15799 of Medoff *et al.*,⁵ or the sequence alignment of SEQ ID NO:3 with GenEmbl accession M30142 of Caras *et al.*^{6,7}

3. It is my opinion that the cited references neither provide motivation to make the claimed compositions, nor provide a reasonable expectation of success in making and using the recited compositions for the reasons explained below.

i. The references do not provide motivation to make the claimed compositions

We previously argued that Powell *et al.* failed to confer permissiveness to CHO and RD cell transfected with DAF and concluded that enteroviruses "can indeed use alternative cellular receptors" different from DAF. Since it is unknown whether enteroviruses use DAF or another unknown receptor for binding to BGМК cells, then there is no motivation to transfect BGМК with DAF (as opposed to transfection with some other unknown receptor).

The Examiner responded that "While Powell *et al.* increase the susceptibility in mouse cells, increased permissiveness to enterovirus infection in a cell expressing a recombinant DAF receptor would only increase susceptibility in a cell that is already susceptible to enterovirus infection, i.e. buffalo green monkey cells. This is due to the established fact in the art that Buffalo green monkey cells are already susceptible to enterovirus infection."⁸

However, The Examiner's assumption is factually wrong because, my experiments that are described in the Specification show that DAF did not alter either sensitivity to enterovirus, or permissiveness to Echo-6 or -11 when DAF was used to transfect H292 cells, even though H292 cells are "already susceptible to enterovirus infection."

For example, the Specification states that:

⁴ Spiller *et al.* (2000) J. Infectious Diseases 181:340-343.

⁵ Medoff *et al.* (1987) PNAS 84(7):2007-2011.

⁶ Caras *et al.* (1987) Nature 325-(6104):545-549.

⁷ Paper No. 17, paragraph bridging pages 6 and 7.

⁸ (Emphasis added) Paper No. 17, page 4, bottom third.

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"The properties and advantages of the invention's transgenic BGMK cells were surprising in view of contrary data disclosed herein when using transgenic H292 cells. In particular, data disclosed herein demonstrates that, whereas transfection of BGMK cells with vectors that express human decay accelerating factor increased both the sensitivity and permissiveness of BGMK cells to enteroviruses, in contrast, no increase in sensitivity to enteroviruses was observed when the same vectors were used to transfect H292 cells."⁹

My results that are disclosed in the Specification using both laboratory isolates¹⁰ and clinical samples¹¹ demonstrate:

"...that transfection of additional copies of the hDAF gene into the H292 cells which express hDAF did not increase or decrease the cells' sensitivity for the detection of laboratory strains of enteroviruses. These results are in direct contrast to those obtained with BGMK-hDAF cells shown in Example 2 *supra*."¹² "In other words, transfection of additional copies of the hDAF gene into H292 cells had no effect on the sensitivity of detection of enteroviruses from in clinical specimens by these cells."¹³

Thus, my results were "surprising" in the face of Scholl *et al.*'s and Powell *et al.*'s disclosure.

Importantly, also, nothing in the art teaches that DAF will confer a new property on the cells i.e., the recited property of selective permissiveness to echovirus-6 and/or echovirus-11 (claim 2 and 4). Nothing in the references suggests that wild type BGMK cells, which are not susceptible to either echovirus-6 or -11, would become susceptible to these particular strains when BGMK cells are transfected to express DAF.¹⁴

⁹ (Emphasis added) Specification, page 6, lines 11-17.

¹⁰ Example 5 beginning on page 50 of the Specification.

¹¹ Example 6 beginning on page 54 of the Specification.

¹² (Emphasis added) Specification, page 54, lines 6-9.

¹³ (Emphasis added) Specification, page 55, lines 1-4.

¹⁴ The Specification confirms that "For echovirus-6 and echovirus-11, BGMK cells failed to detect these two viruses. Importantly, in contrast, BGMK-hDAF cells detected highly diluted virus by day 1."

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II. A reasonable expectation of success in making and using the recited compositions is not established

We previously stated that the specification's data shows that transfecting H292 cells with DAF "had no effect on the sensitivity of detection of enteroviruses..." Therefore, it is unpredictable whether increasing the number of copies of DAF in a cell (e.g., BGMK) that is already susceptible to enterovirus would increase sensitivity to the enterovirus.

The Examiner responded that "These results are not clearly unexpected due to the fact that it is established in the art that human tissue culture cells, i.e. H292, are not as susceptible to enterovirus infection compared with buffalo green monkey cells."¹⁵ In support, the Examiner stated that Melnick "clearly shows that human cell lines are not as susceptible to enterovirus infection compared to monkey kidney cell culture, see Table 2."

However, the Examiner's position is contradicted for at least the following six reasons. First, Melnick does not support the Examiner's position that "H292, are not as susceptible to enterovirus infection compared with buffalo green monkey cells" because Melnick does not make a direct comparison between the specific cell types of H292 (that we used in our investigation) and the invention's BGMK cells. Rather, Melnick's makes a generalized statement by comparing the cytopathic effect of enteroviruses in "monkey kidney" and "human" tissue cultures (Table 2 of Melnick).

Second, Melnick contradicts the Examiner's position by teaching away from using BGMK cells in favor of using human and primary monkey cells for echovirus detection because it says that BGMK cells are not as sensitive to echoviruses as human cells. For example, Melnick says that "...the [BGMK] line may have limitations in sensitivity for routine isolation of a variety of echovirus types, as compared with primary rhesus monkey kidney and human fetal diploid kidney cells."¹⁶

Third, the Specification's data contradicts the Examiner's position that "H292, are not as susceptible to enterovirus infection compared with buffalo green monkey cells" by showing that transgenic H292 cells had the same or better sensitivity as transgenic BGMK cells when infected with Echovirus-4 (days 1-3) and Echovirus-9 (days 2-3) (Table 2 on page 41, and

¹⁵ (Emphasis added) Paper No. 17, page 6, second full paragraph.

¹⁶ Melnick et al., page 661, column 1, second full paragraph.

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Table 6 on page 51), as well as with Coxsackievirus B1 (days 1 and 3; Table 3 page 44 and Table 7 on page 53).

Fourth, Hierholzer *et al.* contradicts the Examiner's statement that "H292, are not as susceptible to enterovirus infection compared with buffalo green monkey cells," because Hierholzer shows that H292 cells supported enterovirus replication to the same extent as other preferred cell cultures. Hierholzer *et al.* observed "that (i) viruses replicated at about the same rate in H292 cells as in their usually preferred cell cultures ..., (ii) virus titer in H292 cells were comparable to those obtained in other, more traditionally preferred cells, and (iii) antigen titers ... were also comparable to those found in supernatant fluids from other cell lines."¹⁷

Fifth, Dagan & Menegus contradicts the Examiner's statement that "H292, are not as susceptible to enterovirus infection compared with buffalo green monkey cells," because this reference (which is co-authored by Dr. Menegus, who is authoritative on enteroviruses) shows that BGMK cells are not as susceptible as human cell lines RD and HEL to enteroviruses.¹⁸

Sixth, the Examiner ignored Applicant's remarks that Powell does not provide a reasonable expectation of success, since Powell failed to confer permissiveness and/or increase sensitivity to enterovirus when transecting RD and CHO cells with DAF. Powell's failure is relevant because it shows that DAF is sufficient to confer permissiveness to enteroviruses in some cases but not others, and there is no basis or guidance in any of the cited references for predicting when success is to be expected.

4. Based on my expertise in the relevant art, and in view of the above discussion, it is my opinion that one of ordinary skill in the art would not have been motivated to make the claimed compositions, nor provided with a reasonable expectation of success in making and using the recited compositions.

¹⁷ Hierholzer *et al.* page 1509, second column, second paragraph; and Table 1, page 1507.

¹⁸ Dagan & Menegus, Figure 1, on page 223.

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5. I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under § 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issuing therefrom.

Dated: March 1st 2004

By: 

Dr. Yung T. Huang

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TAB 1

Revised:Mar.,03

CURRICULUM VITAE

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PHONE: (216) 844-8611 (Office)
(216) 291-2751 (Home)

SOC. SEC.# 535-58-5246

NATIONALITY: U.S.A.

EDUCATION:

BS Degree (1963)	Tunghai University, Department of Biology, Taichung, Taiwan.
MS Degree (1978)	University of North Carolina, Department of Bacteriology and Immunology, Chapel Hill, N.C. 27514
PhD Degree (1983)	University of North Carolina, Department of Microbiology and Immunology, Chapel Hill, N.C. 27514
Postdoctoral (1983)	Dept. of Microbiology and Immunology, University of North Carolina, Chapel Hill, N.C. 27514

PROFESSIONAL APPOINTMENTS:

1984 - present:	Assistant to Associate Professor, Department of Pathology, Case Western Reserve University and Director of Clinical Virology/Serology, Department of Pathology, University Hospitals of Cleveland, Cleveland, Ohio 44106
1979-Jan. 1980:	Research analyst and supervisor, Tissue Culture Facility, Cancer Research Center, University of North Carolina, Chapel Hill, N.C. 27514
1972-1978:	Supervisor, Virology Laboratory, Clinical Microbiology Laboratory, Department of Hospital Laboratories, North Carolina Memorial Hospital, Chapel Hill, N.C. 27514
1970-1971:	Medical laboratory supervisor, U.S. Naval Medical Research Unit No. 2, Taipei, Taiwan
1969-1970:	Special trainee and research technician. School of Public Health,

University of Washington, Seattle, Washington

1964-1969: Senior medical laboratory technician. U.S. Naval Medical Research Unit
No. 2, Taipei, Taiwan

MILITARY SERVICE:

1963-1964: Medical officer in Army (Taiwan).

PROFESSIONAL SOCIETIES:

Member, American Society of Microbiology
Member, American Society of Virology
Member, Pan American Society for Clinical Virology
Member, American Association for the Advancement of Science
Member, American Society of Pathology

RESEARCH SUPPORT:

Biomedical Support Grant 1985-1986 (\$9,883)

Cystic Fibrosis Foundation, Rainbow Chapter 1987-1988 (\$18,546)

National Institute of Health, SCOR: Chronic Diseases of the Airways, 12/1/91 - 11/30/96,
Principal Investigator, E. R. McFadden

Project Title: Interactions of neutrophils with airway epithelial cells *in vitro*, T.D.C. \$569,000
Principal Investigator, Pamela B. Davis, M.D.
Co-investigator with 10% effort

Project Title: Stimulation and evaluation of mucosal immunity to respiratory pathogens, T.D.C.
\$929,734
Principal Investigator, John G. Nedrud, Ph.D.
Co-principal investigator, Michael E. Lamm, M.D.
Co-investigator with 15% effort

National Institute of Health, Program project: Mucosal Immunity and Infection, 9/1/95 -
8/30/98,
Project Director, M. E. Lamm
Principle Investigator with 30% effort
Project Title: Intracellular neutralization of SIV/HIV, T.D.C. \$144,000

Grant from Diagnostic Hybrids INC, Athens, Ohio.
Project Title: Investigation of Turbo for viral diagnosis
1/1/97 - 12/31/98 T.D.C. \$40,000

National Institute of Health, Project: Studies on secretory Immunoglobulin 01-01-98 to 12-31-02
Principal Investigator: Michael E. Lamm
Co-investigator with 15% effort.

National Institute of Health, Project: FAS-Ligand Mediated Immunity to HSV.
07-01-99 to 06-30-2004
Principle Investigator: David Kaplan
Co-investigator with 4% effort

National Institute of Health,
Program project: Mucosal immunity and infection 10-1-00 to 9-30-05
Project 3: Principal Investigator
Project title: IgA in resisting HIV. TDC \$754,179.00

Ohio Dept. of Development (Technology Action Fund).
Fund period: 6-03 to 5-05
Principle investigator
Project title: Genetically Engineered cell lines for Improving Influenza Vaccine Production and
Rapid Respiratory virus Detection.
Total funding: \$401,390.00

BOARD CERTIFICATION:

American Board of Bioanalysis: Certified High-Complexity Clinical Laboratory Director

OTHER:

First patent granted on April 6, 1999.
Second patent granted on January 2, 2001.
Third patent granted on August 28, 2001.
Fourth patent granted on October 23, 2001
Fifth patent granted on April 23, 2002
Sixth patent granted on April 25, 2002
Seventh patent granted on June 18, 2002
Eighth patent granted on December 17, 2002
Ninth patent granted on August 26, 2003
All licensed to Diagnostic Hybrids Inc. Athens, Ohio.

COMMITTEE APPOINTMENTS:

Hospital Laboratory Quality Improvement Committee, 1996 - 1998
Pediatric Infection Control Committee, 1990 - Present
Pathology Department Library Committee, 1995-Present

PUBLICATIONS:

ORIGINAL PAPERS:

1. Lee GCY, Funk GA, Chen ST, Huang YT, and Wei HY. An outbreak of respiratory syncytial virus infection in an infant nursery. J Formosan Med Assoc 72:39-46, 1973
2. Huang YT, Huang ES, and Pagano JS. Specific antisera to cytomegaloviruses. J Immunol 112:528, 1974
3. Lee GCY, Huang YT, and Chang LC. Preliminary study of attenuated Japanese Encephalitis Virus. J Formosan Med Assoc 74:606-12, 1974
4. Hutt LM, Huang YT, Dascomb HE, and Pagano JS. Cytotoxicity of leukocytes during mononucleosis. J Immunol 115:243-48, 1975
5. Huang ES, Kilpatrick BA, Huang YT and Pagano JS. Detection of human cytomegalovirus and analysis of strain variation. Yale J Biol and Med 49:29-43, 1976
6. Pagano JS, Huang CH, and Huang YT. Epstein-Barr virus genome in infectious mononucleosis. Nature 263:787-89, 1976
7. Lemon SM, Hutt LM, and Huang YT. Cytotoxicity of circulating leukocytes in cytomegalovirus mononucleosis. Clin Immunol and Immunopathology 8:513-19, 1977
8. Lemon SM, Hutt LM, Huang YT, Blum JE and Pagano JS. Simultaneous infection with multiple Herpes viruses. Amer J Med 66:270-76, 1979
9. Huang YT and Wertz G. Respiratory syncytial virus is a negative stranded RNA virus that code for at least seven complementary message RNA species. J Virol 43:150-57, 1982
10. Huang YT and Wertz G. Coding assignments to the six mRNAs of respiratory syncytial virus. J Virol 46:667-72, 1983
11. Collins PL, Huang YT and Wertz GW. Identification of a tenth mRNA of respiratory syncytial virus and assignment of polypeptides to the ten viral genes. J Virol 49:572-78, 1984
12. Collins PL, Huang YT and Wertz GW. Nucleotide sequence of the gene encoding the fusion (F) glycoprotein of human respiratory syncytial virus. Proc Natl Acad Sci 81:7683-87, 1984
13. Huang YT, Collins PL and Wertz GW. Characterization of the 10 proteins of human respiratory syncytial virus: Identification of a fourth envelope associated protein. Virus

Research 2:157-73, 1985

14. Wertz GW, Collins PL, Huang YT, Gruber C, Levine S and Ball A. The G protein of human respiratory syncytial virus constitutes a novel type of viral membrane protein. Proc Natl Acad Sci 82:4075-79, 1985
15. Midulla F, Huang YT, Gilbert IA, Cirino NM, McFadden ER, Jr and Panuska JR. Respiratory syncytial virus infection of human cord and adult blood monocytes and alveolar macrophages. Am Rev Respir Dis 140:771-77, 1989
16. Weaver MG, Abdul-Karim FW, Dale G, Sorensen K and Huang YT. Detection and localization of human papillomavirus in penile condylomas and squamous cell carcinomas using in situ hybridization with biotinylated DNA viral probes. Modern Pathol 2:94-100, 1989
17. Panuska JR, Cirino NM, Midulla F, Despot JE, McFadden ER, and Huang YT. Productive infection of isolated human alveolar macrophages by respiratory syncytial virus. J Clin Invest 86:113-19, 1990
18. Panuska JR, Midulla F, Cirino NM, Villani A, Gilbert IA, McFadden ER, Huang YT. Human mononuclear phagocytes infected with respiratory syncytial virus have altered production of tumor necrosis factor-alpha and prostaglandin E₂. Am J Physiol 259:L396-L402, 1990
19. Weaver MG, Abdul-Karim FW, Dale G, Sorensen K, Huang YT. Outcome in mild and moderate cervical dysplasias related to the presence of specific human papillomavirus types: a retrospective study using in situ hybridization with biotinylated DNA viral probes. Modern Pathol 3:679-83, 1990
20. Smith MC, Creutz C, Huang YT. Detection of respiratory syncytial virus in nasopharyngeal secretions by shell viral techniques. J Clin Microbiol 3:463-65, 1991
21. Stark JM, Fatemi SH, Amini SB, Huang YT. Occurrence of respiratory syncytial virus subtypes in hospitalized children in Cleveland, Ohio from 1985-1988. Pediatric Pulmonology 11:98-102, 1991
22. Stark JM, Huang YT, Davis PB. Infection of cultured human tracheal epithelial cells by human parainfluenza virus types 2 & 3. J Virological Methods 3:31-46, 1991
23. Tosi MF, Stark JM, Hamedani A, Smith CW, Gruenert DC, Huang YT. Intracellular adhesion Molecule I (ICAM-1)-dependent and ICAM-1-independent adhesive interactions between polymorpho-nuclear leukocytes and human airway epithelial cells infected with parainfluenza virus type 2. J Immunol 149:3345-49, 1992
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- hybridization. J Virol Method 41:1-8, 1993
25. Huang YT, Romito RR, Bishnn PD, Banerjee AK. Characterization of the in vitro system for the synthesis of mRNA from human respiratory syncytis/virus. Virology 193:862-67, 1993
 26. Huang R, Romito R, Panin M, Huang YT. Effect of 6-diazo-5-oxo-L-norleucine on replication of human respiratory syncytial virus. Antiviral Research 25:269-79, 1994
 27. Sieg S, Muno-Cacho C, Robertson S, Huang YT, Kaplan D. Immunoregulation of human T lymphocytes by parainfluenza virus type 3. Proc Natl Acad Sci USA, 91:6293-97, 1994
 28. Hemingway BR, Yang Y, Panin M, Huang YT, Galinski MS. Role of basic residues in the proteolytic activation of Sendai virus fusion glycoprotein. Virus Research 36:15-35, 1995
 29. Huang YT, Romito RR, Panin M. Characterization of human parainfluenza virus type 2 RNAs in infected cells and by in vitro synthesis. Virus Research 35:181-92, 1995
 30. Kiyono H, Miller C, Lu Y, Lehner T, Cranage M, Huang YT, Kawabata S, Marthas M, Roberts B, Nedrud J, Lamm M, Bergmeier L, Brooks R, Tao L, McGhee J. The common mucosal immune system for the reproductive tract: basic principle applied towards an AIDS vaccine. Advanced Drug Delivery Reviews 18:23-51, 1995
 31. Pappin A, Grissom M, Huang YT, Yomtovian R. Beyond the seven day limit: Stability of cytomegalovirus antibodies in plasma during prolonged storage of blood components. Clin and Diag Lab Immun 2:25-29, 1995
 32. Sieg S, Xia L, Huang YT, Kaplan D. Specific inhibition of granzyme B by parainfluenza virus type 3. J. Virol 69:3538-41, 1995
 33. Sokhandan M, McFadden ER, Huang YT, Mazanec MB. The contribution of respiratory viruses to severe exacerbations of asthma in adults. Chest 107:1570-74, 1995
 34. Hite S, Huang YT. Microwave-accelerated direct immunofluorescent staining for RSV and influenza virus. J Clinical Micro 34:1819-20, 1996
 35. Mazanec MB, Huang YT, Pimplikar SW, Lamm ME. Mechanisms of inactivation of respiratory viruses by IgA, including intraepithelial neutralization. Seminar in Virol 7:285-92, 1996
 36. Sieg S, King C, Huang YT, Kaplan D. The role of interleukin-10 in the inhibition of T-cell proliferation and apoptosis mediated by parainfluenza virus type 3. J Virol 70:4845-48, 1996

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38. Sieg S, Huang YT, and Kaplan D. Viral regulation of CD95 expression and apoptosis in T lymphocytes. J. Immunol. 159:1192-1199, 1997
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40. Huang YT, Miller CJ, Wong V, Fujioka H, Nedrud J, Lamm M L. Replication and budding of Simian immunodeficiency virus in polarized epithelial cells. Virology 257:24-34, 1999
41. Huang YT, Turchek B. Mink lung cells and mixed mink lung and A549 cells for rapid detection of influenza virus and other respiratory viruses. J. Clinical Microbiol. 38: 422-423, 2000
42. Huang YT, Hite S, Duane V, Yam P, and Jollick JA. Application of mixed cell lines for the detection of viruses from clinical specimens. Clinical Microbiology Newsletter. 22: 89-92, 2000
43. Huang YT, Hite S, Duane V, and Yan H. CV-1 and MRC-5 mixed cells for simultaneous detection of herpes simplex viruses and varicella zoster virus in skin lesions. J Clinical Virol. 24: 37-43, 2002
44. Huang YT, Yam P, Yan H, and Sun Y. Engineering BGMK cells for sensitive and rapid detection of enteroviruses. J Clinical Microbiol. 40: 366-371, 2002
45. Huang YT, Yan H, Sun Y, Jollick JA, and Baird H. Cryopreserved cell monolayers for rapid detection of herpes simplex virus and influenza virus. J Clinical Microbiol. 40: 4301-4303, 2002
46. Yan H, Lamm ME, Bjorling E and Huang YT. Multiple functions of immunoglobulin A in mucosal defense against viruses: an invitro measles virus model. J Virol. 76: 10972-10979, 2002

ABSTRACTS AND PAPERS IN BOOKS:

1. Huang YT, Davis NL and Wertz GW. Separation and characterization of the RNAs of human respiratory syncytial virus. IN: Replication of Negative Stranded RNA Viruses. Bishop, D.H.L. and Company, R.W., Eds. Elsevier, p. 531-36, 1981

2. Huang YT and Wertz G. Analysis of the RNAs of human respiratory syncytial virus. Abstracts American Soc. Microbiology T-91, 1981
3. Huang YT and Wertz G. Structural and functional analysis of the RNAs of human respiratory syncytial virus infected cells. Abstracts, Fifth International Congress of Virology, pp. 400, 1981
4. Huang YT, Collins PL and Wertz GW. Identification of a new envelope-associated protein of human respiratory syncytial virus. In: Non-segmented negative strand viruses. Bishop D.H.L. and Company, R.W. Eds., pg. 365-68, 1984
5. Huang YT, Collins PL and Wertz GW. The polypeptides of human respiratory syncytial virus. Abstracts American Soc. Microbiology T-27, 1984
6. Rosenthal GE, Sanford S, Huang YT, Heggie AD and Landefeld CS. Validation of a rule estimating the risk of chlamydial infection in women. Presented to American Federation of Clinical Research, April 1988
7. Heggie AD, Roessmann U and Huang YT. Host factors in coxsackie virus B-2 disease. Presented to annual meeting of American Pediatric Society, Washington D.C., May 1988
8. Weaver MG, Abdul-Karim FW, Dale G, Sorensen K, and Huang YT. Detection and localization of human papillomavirus in penile condylomas and squamous cell carcinomas using in situ hybridization with biotinylated DNA viral probes. Presented to United States and Canadian Academy of Pathology meeting, September 1988
9. Midulla F, Panuska JR, Huang YT, Gilbert I, Cirino N, and McFadden ER. Respiratory syncytial virus infection of human alveolar macrophages: Effects of differentiation lipopolysaccharide activation. Presented to XIII International Congress of Allergy and Clinical Immunology, October 1988
10. Stark JM, Huang YT, and Davis PB. Parainfluenza and respiratory syncytial virus infection of cultured human epithelial cells - A possible model for lower respiratory tract disease. Presented to the annual meeting of American Thoracic Society, November 1988
11. Midulla F, Huang YT, Gilbert I, Cirino N, McFadden ER and Panuska J. Respiratory syncytial virus infection of human mononuclear phagocytes. Presented to annual meeting of American Thoracic Society, January 1989
12. Tosi M, Stark JM, Hamedani A, Smith CW, Gruenert DC and Huang YT. Increased adhesion by neutrophils to human tracheal epithelial cells infected with parainfluenza virus type 2: role of epithelial ICAM-1 and neutrophil CD11/CD18 adhesions.

- Presented to annual meeting of the American Pediatric Society, April 1991
13. Sokhandan M, McFadden ER, Huang YT, Mazanec MB. The contribution of respiratory viruses to exacerbations of asthma in adults. Presented to annual meeting of the American Thoracic Society, May 1991
 14. Heggie AD, Scholl DR and Huang YT. Rapid detection of herpes simplex viruses in culture by in situ hybridization. Presented to Annual Clinical Virology Symposium, April 1992
 15. Huang YT, Romito RR, De BP, Banerjee AK. In vitro RNA synthesis of human respiratory syncytial virus. Submitted to annual meeting of American Society of Virology, July 1992
 16. Panin M, Liang XP, Nedrud JG, Huang YT. Immune response in mice upon oral immunization with attenuated salmonella typhimurium expressing Np of Sendai virus. Submitted to annual meeting of the American Society of Virology, July 1992
 17. Galinski MS, Hemingway BR, Yang Y, Panin M, Huang YT. Role of basic residues in the proteolytic activation of Sendai virus fusion glycoprotein. Submitted to the annual meeting of American Society of Virology, July 1993
 18. Pappin A, Grissom M, Huang YT, Yomtovian R. Beyond the seven day limit: Stability of cytomegalovirus antibodies in plasma during prolonged red blood cell storage. Submitted to annual meeting of American Society of Clinical Pathology, August 1994
 19. Hite S, Huang YT. Microwave-accelerated direct immunofluorescent staining for RSV and influenza virus. Submitted to the annual meeting of the North American Society for Clinical Virology, April 1995
 20. Huang YT, Miller C, Nedrud J, Lamm M. Replication and budding of SIV in polarized epithelial cells. Submitted to the annual Symposium on Nonhuman Primate Models for AIDS, November 1995
 21. Huang YT, Dul J, Brown J, Scholl D. Shell vial detection of CMV using mink lung cells enhanced by treatment with CMV TurboTreat™: A new viral culture medium. Submitted to the Annual Meeting of the North American Society for Clinical Virology, April 1997
 22. Turchek B, Huang YT. Typing of HSV isolates detected by Elvis HSV test. Submitted to the Annual Meeting of the North American Society for Clinical Virology, April 1997
 23. Friedland RP, Lerner AJ, Smith AL, Huang YT, Siedlak SL, Perry G. Latent infection with cytomegalovirus in Alzheimer's Disease (AD): Serum antibody and immunocytochemical studies. Submitted to the American Society for Neurology, 1997

24. Huang YT, Turchek B. Sensitive cell lines for detection of respiratory viruses from clinical specimens: comparison of mink lung and NCIH292 to primary rhesus monkey kidney and Hep-2 cells. Presented at Annual Meeting of the North American Society for Clinical Virology, April 1998
25. Huang YT, Hite S, Duane V. CV-1/MRC-5 mixed cells for detection of herpes simplex viruses: comparison with primary rabbit kidney and mink lung cells. Presented at Annual Meeting of the North American Society for Clinical Virology, May 1999
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